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***ACTINEMYS MARMORATA* (Northern Pond Turtle). DIET.** *Actinemys marmorata* has been described as a dietary generalist, exploiting a broad food niche that is primarily aquatic, but sometimes includes terrestrial foods (Bury 1986. J. Herpetol. 20:515–521). Much of the diet of adults is plant-based, the bulk of which is filamentous algae. Stomach contents analyses have revealed that these turtles are also opportunistic predators of live fish and larval and adult anurans (Bury 1986, *op. cit.*), although predation events are rarely witnessed in the wild (Major 1991. Auk 108:190–195; Van Vuren 2001. Am. Midl. Nat. 145:94–100). However, a seasonal abundance of protein-rich prey appears each spring in the form of amphibian eggs (and unhatched larvae). Biphasic amphibians whose life history includes aquatic larvae must use aquatic habitats to reproduce, and the egg-laying of aquatic amphibians is often concentrated within rather short time periods and offers immobile, easily-obtained food for an opportunistic predator. Here, I report on a direct observation of an *A. marmorata* feeding on amphibian eggs.

My observation occurred in a small, narrow pool in an intermittent stream in Sonoma County, California, USA, on the afternoon of 18 May 2018. Copeland Creek originates on the Mitsui Ranch, at the top of Sonoma Mountain, and meanders through a riparian strip comprised primarily of California Bay-laurel (*Umbellularia californica*), Oregon Oak (*Quercus garryana*), Coast Live Oak (*Quercus agrifolia*), and willows (*Salix* spp.), surrounded by open grasslands. While surveying for native amphibians in an open-canopy stretch of the creek, I noticed a small area of Quaking Grass (*Glyceria leptostachya*) and sedges (*Carex* spp.) in the water near the shear bank that edges a small pool. Nothing was visible on the surface, so I stepped cautiously forward to the edge to get a closer look. Half a meter below my feet, the bank was slightly undercut and I could see the head of a Northern Pond Turtle moving just under the surface. The turtle's head was repeatedly lunging forward toward some vegetation, but it wasn't coming away with any pieces; the vegetation kept springing back each time the turtle pulled away. When I raised my 8 × 42 binoculars, I could see that the turtle was biting off portions of a gelatinous mass that was attached the grass stem. After a moment, I recognized the curved arc of a developing amphibian embryo at the approximate developmental stage of 20–25 (Gosner 1962. Herpetologica 16:183–190), but there was not enough of the embryo mass remaining to visually determine the species. However, as I watched the turtle (a male, ca. 140 mm carapace length) over the next 4 min, it fully consumed three embryo masses of the Pacific Chorus Frog (*Hyliola regilla*). The turtle never surfaced while I watched, and though there were many embryo masses in the pool, each at various stages of development, the turtle moved past the less developed ones and appeared to target those in the latter stages. Two of the masses it consumed were attached obliquely to grass stems and the turtle consumed these quickly, but a third mass was wrapped around a grass stem, and the turtle required more time and effort with that one. After consuming this difficult mass, the turtle surfaced. It spotted me and immediately dove under a rock in the deepest part of the pool, ending my observation.

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***BATAGUR TRIVITTATA* (Burmese Roofed Turtle). SEXUAL SIZE DIMORPHISM.** *Batagur trivittata* is a critically endangered, highly aquatic turtle endemic to the large rivers of Myanmar (Ernst and Barbour. 1989. Turtles of the World. Smithsonian Institution Press, Washington, D.C. 313 pp.; Stanford et al. 2018. Turtles in Trouble: The World's 25+ Most Endangered Tortoises and Freshwater Turtles - 2018. IUCN Tortoise and Freshwater Turtle Specialist Group, Ojai, California. 79 pp.). *Batagur trivittata* was feared extinct (Bhupathy et al. 2000. Chelon. Res. Monogr. 2:156–164) until it was "rediscovered" in the early 2000s (Platt et al. 2005. Chelon. Conserv. Biol. 4:942–948; Kuchling et al. 2006. Oryx 40:176–182), and intense *in-* and *ex-situ* conservation efforts are now focused on population recovery (Platt and Platt 2018. Turtle Survival 2018:41–48). Although fewer than 10 reproductive females survive in the wild (Çilingir et al. 2018. Conserv. Biol. 31:1469–1476), the captive population is approaching 900 and thus, *B. trivittata* is no longer in eminent danger of biological extinction. Conservation efforts notwithstanding, basic natural history information on *B. trivittata* remains sparse. Smith (1931. The Fauna of British India, including Ceylon and Burma. Vol. 1. Loricata and Testudines. Taylor and Francis, London, UK. 185 pp.) stated that female *B. trivittata* (carapace length [CL] to 580 mm) are considerably larger than males (CL to 460 mm), but otherwise little is known about sexual size dimorphism in this species. We here quantify sexual size dimorphism in a large series (N = 179) of *B. trivittata* and comment on the adaptive significance of our findings.

Our sample of turtles was sourced from captive-breeding colonies at Htamanthi Wildlife Sanctuary (N = 70), Lawkanandar Wildlife Sanctuary (N = 20), Wildlife Reserves Singapore (N = 22), Yadanabon Zoological Gardens (N = 41), and Yangon Zoological Gardens (N = 20). Five turtles held at Yadanabon Zoological Gardens were obtained as adults from pagoda ponds or confiscated from traders and fishermen; the remaining turtles in our sample were reared in captivity after being hatched from eggs deposited by captive females or collected from nests along the Chindwin River. All turtles used in our study were ≥ 10 years-old, and breeding coloration was evident in males during the reproductive season. We determined the sex of each turtle based on dimorphic shell coloration and tail morphology (Platt et al. 2017. Herpetol. Rev. 48:616–618), and measured (to nearest 1.0 mm) the mid-line CL (Method D; Iverson and Lewis 2018. Herpetol. Rev. 49:453–460) using tree calipers. In addition to captive turtles, we also included measurements of six intact shells of adult female *B. trivittata* examined at villages along the Dokhtawady and Chindwin rivers (Platt et al. 2005, *op. cit.*; Platt et al. 2018. Nat. Hist. Bull. Siam Soc. 63:67–114) in our analysis.

We used a Student's t-test to test the one-tailed hypothesis that CL of adult female *B. trivittata* was significantly greater than that of adult males (e.g., Platt et al. 2008. Chelon. Conserv. Biol. 7:195–204). We then quantified the degree of size dimorphism (defined as a statistically significant difference in mean length or mass of organisms from the same population) between the sexes with a compressed sexual size dimorphism index (SDI;